



Evaluation of *In Vitro* anthelmintic activity of *Cymbopogon citratus* (lemon grass) extract

Sikandar Khan Sherwani^{1*}, Muhammad Mumtaz Khan², Muhammad Uzair Khan³, Muhammad Ajmal Shah⁴ and Shahana Urooj Kazmi⁵

1, Department of Microbiology, Federal Urdu University of Arts, Science and Technology, Karachi, Pakistan

2, Department of Biotechnology, University of Karachi, Karachi, Pakistan

3, Department of Microbiology, Kohat University of Science and Technology, KPK, Pakistan

4, Department of Pharmacognosy, Federal Urdu University of Arts, Science and Technology, Karachi, Pakistan

5, Department of Microbiology, University of Karachi, Karachi, Pakistan

Abstract

Lemon grass a popular aromatic plant which is commonly used as a substitute of green tea has a tremendous phytomedicinal potential. Like other tea plants it also having antimicrobial as well as antiprotozoal potential. Main objective of the present study was to find out the anthelmintic potential of aqueous extract of *Cymbopogon citratus* (lemon grass) by using a model worm i.e. *Pheretima posthuma* (earthworm). In this bioassay, three different concentrations i.e. 25, 50 and 1000mg/ml of above mentioned extract were tested in bioassay by noting time of paralysis and time of death of worms in minutes. Piperazine citrate was used as a standard reference compound along with normal saline as control in the same concentrations. The results indicated that lemon grass crude extract possessed anthelmintic activity in dose dependent manner.

Key-Words: Helminthes, *Cymbopogon citratus*, Lemon grass, *Pheretima posthuma*, Anthelmintic

Introduction

Since the beginning of human history, plants have been in practice as folk medicine. All natural plants are rich source of medicinal agents, commonly in traditional medicine (Bonjar *et al.*, 2004). One or more parts of medicinal plants having substances that can be useful for the therapeutic purpose (Rios, 2005). *Cymbopogon citratus* belongs to the Poaceae family which is monocotyledonous aromatic perennial having slender sharp edged leaves and pointed apex commonly inhabitant to tropical Asia. In west India and Pakistan, *C. citratus* is also known as Guatemala, lemongrass (Ernst, 2008). The extract of *C. citratus* having different compounds where as the major component is citral approximately 65-85%. Additionally, lemongrass extracts having small quantity of geranylacetate, monoterpene olefins and geraniol (Fair & Kormas, 2008). Lemon grass are being used as the therapeutic agent for the treatment of gastrointestinal disturbances, nervous, hypertension and fever were used.

*** Corresponding Author**

Email: sikander_biology@hotmail.com

Contact: +923245189042

The extracts of *C. citratus* are also used in elephantiasis, coughs, flu, headache, gingivitis, malaria, leprosy, ophthalmia, vascular disorders and pneumonia. Several studies of lemongrass extracts have been reported on its antifungal and antibacterial activities (Onawunmi *et al.*, 1989; Wannissorn *et al.*, 1996; Schuck., 2001; Paranagama; 2003; Wannissorn *et al.*, 2005). Moreover, antiviral potential especially against Herpes simplex virus were also reported (Minami *et al.*, 2003). Gore *et al.* in 2010 reported the anti helminthes activities of lemongrass (Gore *et al.*, 2010) Helminths are most common agents spread human infections frequently in developing countries and contribute by increasing the global burden of diseases actively (Hotez *et al.*, 2008). World Health Organization (WHO) estimated 2 billion people with helminthic infection and this population was suggested to get medicine to expel and kill the helminthes from human body (WHO, 2012). The purpose of present study is to investigate the anthelmintic activity of lemongrass against earthworm.

Material and Methods

Preparation of extract

The lemon grass tea was purchased locally from the retail market in Karachi-Pakistan in reasonable price and brought to the laboratory of Federal Urdu University of Arts, science and Technology (FUUAST)-Karachi-Pakistan. The extract was prepared in 5% concentration (5 grams of weighted lemon grass tea added in 100 ml of distilled water). The aqueous extract was prepared by boiling method of tea in water bath by constant agitation of for 15 minutes. After extract preparation, coarse suspended particles of tea were removed by passing through strainer and later by passing via 0.22um filter (Sherwani *et al.*, 2013). The extract was stored in refrigerator in small vials as aliquots for further use.

Selection of worms

The anthelmintic activity was done by following the method of (Ajaiyeoba *et al.*, 2001) with slight modifications. The assay was carried out on adult earthworm (*Pheretima posthuma*) owing to its resemblance in terms of anatomy and physiology with the intestinal roundworm parasite of human beings (Vigar, 1984; Sherwani *et al.*, 2013; Thorn, 1977; Chaterjee 1967; Vidyarthi 1967). This worm has been a choice as a model for researchers to investigate anthelmintic activity of natural plant product is its easy availability (Szewezuk *et al.*, 2003; Deore *et al.*, 2009). Because of this reason; earthworms have been used commonly for in vitro evaluation at initial level of anthelmintic bearing natural and synthetic substances (Sollmann, 1918; Jain and Jain., 1972; Shikvar and Kumar., 2003).

Worm Collection and authentication

Earthworms of approximately within the size range of 8 cm (Vidyadhar *et al.*, 2010) were collected from moist garden soil of Federal Urdu University of Arts, science and Technology-Karachi Pakistan with the help of a Zoologist from MRCC-Department-University of Karachi-Pakistan and authenticate the *Pheretima posthuma*. Later, all the worms were washed with saline to remove the soil particles, debris and fecal material (Satish and Ravandra, 2009) and kept them in phosphate buffer saline (PBS) (Eugale and Giday, 2009) till further work.

Anthelmintic assay

The earthworms were divided into three groups having six earthworms. Group first serve as control, receive only normal saline; Group second serve as standard, receive standard piperazine citrate and Group third serve as aqueous extract of different concentration (Dwivedi *et al.*, 2011). All the extracts and the standard solution in autoclaved distilled water were freshly

prepared before conducting the assay (Rajesh *et al.*, 2010). The concentration of control, standard and extract that were used in this assay were 25 mg/ml, 50mg/ml and 100mg/ml. Observations were made for the time taken until the paralysis as well as death of an individual worm occurred (Trapti *et al.*, 2009; Grime *et al.*, 2006). The mean time of paralysis and death was recorded in minutes. The paralysis was declared when the worms were not able to move even in normal saline (Zafar *et al.*, 2001). Death was considered when the worms lost their motility followed with fading away of their body colors (Mali and Mahale, 2008).

Results and Discussion

Helminthes infection like other infections are also the most wide spread infections in humans, affecting seriously a huge population of the world (Kosalge and Frusule, 2009). Moreover, there is a need to pay serious attention to the existing helminthic infections as the majority of infections owing to helminthes are not only injurious to health but also play a role somehow in the development of anemia, pneumonia, undernourishment, eosinophilia and some other secondary complications (Bundy, 1994). Anthelmintic or wormicidal agents are the drugs that expel out parasitic worms (helminthes) from the body by causing paralysis or killing them but these anthelmintic drugs are also now adopting resistance and therefore, no more effective in management of infections (Chartier *et al.*, 2001). The plants are very well known to provide a rich and diverse source of botanical anthelmintic (Coles, 1997). In this study, in earthworms, the aqueous extract of *Cymbopogon citratus* (lemon grass) showed anthelmintic activity. A number of previously conducted report indicated that that *Cymbopogon citratus* has been used against gastrointestinal disturbances and complications (Nakamura *et al.*, 2003). Earthworms belonging to control group showed paralysis time at the concentration of 25mg/ml, 50mg/ml and 100mg/ml i.e. 96.46 ± 0.46 mins, 78.23 ± 0.25 mins and 62.43 ± 0.11 mins while; death time 160.06 ± 0.11 mins, 142.2 ± 0.34 mins and 111.7 ± 0.26 mins respectively. On the other hand, the standard reference compound piperazine citrate showed the time of paralysis time at the concentration of 25mg/ml, 50mg/ml and 100mg/ml i.e. 18.36 ± 0.15 mins, 12 ± 0.00 mins and 08.84 ± 0.03 mins respectively while; death time at 54.36 ± 0.40 mins, 46.46 ± 0.40 mins, 13.6 ± 0.17 mins respectively as indicated in Table 1. In case of aqueous extract of *Cymbopogon. citratus* (lemon grass) group, its paralysis time at the concentration of 25mg/ml, 50mg/ml and 100mg/ml i.e. 44.86 ± 0.41 mins, 22.16 ± 1.06 mins and 10.7 ± 0.43 mins respectively death time at 120.46 ± 0.25 mins, 92 ± 0.51

mins and 80.7 ± 0.51 mins respectively as indicated in Table 2. The activity was observed in dose dependent way in all groups including control, standard drug and extract. From the observations achieved, higher concentration of crude extract of lemon grass showed paralytic effect much earlier and the time to death was shorter for almost all the worms. The results of similar study but conducted on green tea extract at (50mg/ml) showed paralysis within 27.02 min & time of death 47.07 mins while, aqueous extract (100mg/ml) showed paralysis within 21.19 min & time of Death 37.02 mins (Dwivedi *et al.*, 2010). According to mode of action, piperazine citrate causes flaccid paralysis and expel out the worm by peristalsis (Sherwani *et al.*, 2013). Piperazine citrate generates paralysis by increasing the chloride ion conductance of worm muscle membrane resulting in the hyper polarization and reduced excitability that ultimately leads to muscle relaxation thus a worm expels out by the action of peristalsis (Goshwami *et al.*, 2013). Phytochemical analysis of *Cymbopogon citratus* revealed presence tannins (Ewansiha *et al.*, 2012) and considered to inhibit the infective agents (Kolodziej & Kiderlen, 2005). Some other polyphenolic compounds also known to have anthelmintic activity (Bate Smith, 1962).

Conclusion

From the results of this preliminary work, it is concluded that the crude aqueous extract of *Cymbopogon citratus* (lemon grass) showed anthelmintic activity and could be apply as an effective agent in future after further exploration. Studies should be needed in next steps of the undertaken work for understanding the mechanism of action by using in vivo models to figure out the effectiveness and pharmacological rationale of using lemon grass as an anthelmintic drug.

Acknowledgement

The authors are highly thankful to Dr. Uzair Khan, Assistant Professor-Marine Reference Collection (MRCC)-University of Karachi-Karachi-Pakistan for helping in the collection and identification of earthworms from the garden soil and Dr. Kanwal Nazim, Assistant Professor-Marine Reference Collection (MRCC)-University of Karachi-Karachi-Pakistan for providing standard reference compound.

References

1. Ajaiyeoba EO, Onocha PA, Olarenwaju OT. In vitro anthelmintic properties of Buchholzia coriacea and Gynandropsis gynandra extract. Pharm Biol 2001 39:217-20.
2. Bate Smith E.C. The phenolic constituent of plants and their taxonomic significance,

dicotyledons. J Linn Soc Bot 1962; 58:95-103.

3. Bonjar, G.,H.,S & Farrokhi, P.,R. (2004). Antibacillus activity of some plant used in traditional medicine of Iran. Nigerian Journal on National Prod. Med. (8): 34-39.
4. Bundy D A. Immunoepidemiology of intestinal helminthic infection I: The global burden of intestinal nematode disease. Trans Royal Soc Trop Med Hyg 1994; 8: 259-261.
5. Chatterjee KD. Parasitology, Protozoology and Helminthology. 6th ed. Calcutta: In Guha Ray Sree Saraswaty Press Ltd; 1967.
6. Chartier, C., Soubirac, F., Pors, I., Silvestre, A., Hubert, J., Couquet, C., Cabaret, J. (2001). Prevalence of anthelmintic resistance in gastrointestinal nematodes of dairy goats under extensive management conditions in south-western France. J. Helminthol., 75, 325-330.
7. Coles GC. Nematode control practices and anthelmintic resistance on British sheep farms. Vet. Rec. 1997; 141: 91-93.
8. Deore S. L., S.S.Khadabadi, Kamdi K. S., Ingle V. P., Kawalkar N. G. Sawarkar P. S, Patil U. A, Vyas A. In vitro Anthelmintic activity of Cassia tora. International Journal of ChemTech Research. Vol.1, No.2, pp 177-179, 2009
9. Dwivedi G, Bairagi. M, Rawal. D, Rawal. S. Anthelmintic activity of *Myristica fragrans* (Nutmeg) extracts 2011. RJPBCS Volume 2 Issue 2 Page No. 315
10. Eguale. T and Giday. M.. In vitro anthelmintic activity of three medicinal plants against *Haemonchus contortus* 2009; 3 : 1 : 29-34
11. Ewansiha J. U., Garba S. A., Mawak J. D., Oyewole O. A. Antimicrobial Activity of *Cymbopogon Citratus* (Lemon Grass) and It's Phytochemical Properties. Frontiers in Science 2012, 2(6): 214-220
12. Ernst, E. (2008). "Chiropractic: a critical evaluation". Journal on Pain Symptom Management 35 (5): 544-62
13. Fair, J.D & Kormas, C.M. (2008). Journal on Chromatography, 1211(1-2), 49-54.
14. Girme AS, Bhalke RD, Ghogare PB, Tambe VD, Jadhav RS, Nirmal SA: Comparative *in vitro* anthelmintic activity of *Mentha piperita* and *Lanata camara* from western India, Dhaka Univ. J. Pharm. Sci. 5(1-2),2006:5-7
15. Goshwami . D, Rahman. M, Muhit. M and Saiful Islam. Evaluation Of Anthelmintic

- Activity of *Lasia spinosa* Leaves. International Journal of Current Pharmaceutical Research Vol 5, Issue 1, 2013.
16. Hotez, P.J. 2008. Forgotten people and forgotten diseases, the neglected tropical diseases and their impact on global health and development. *ASM Press*. In press
 17. Jain ML, Jain SR. Therapeutic utility of *Ocimum basilicum* var. album. *Planta Med* 1972; 22:66-70
 18. Kolodziej, H & Kiderlen, A.F. (2005). "Antileishmanial activity and immune modulatory effects of tannins and related compounds on *Leishmania* parasitised RAW 264.7 cells". *Phytochemistry* 66 (17): 2056-71.
 19. Kosalge S.B and Fursule R. B. Investigation Of In Vitro Anthelmintic Activity Of *Thespesia Lampas* (Cav.) *Asian Journal of Pharmaceutical and Clinical Research. Volume 2, Issue 2, April- June, 2009*
 20. Minami M., Kita M., Nakaya T., et al. The inhibitory effect of essential oils on Herpes simplex virus type-1 replication *in vitro*. *Microbiol Immunol* 2003; 47: 681-4.
 21. Mali RG and Mahale NB: Evaluation of *Rhynchosia minima* (Linn.) DC leaves for antihelmintic activity, *International Journal of Pharmaceutical Sciences and Nanotechnology*, 1(2), 2008: 191-194.
 22. Nakamura, Y.; Miyamoto. M.; Murakami, A.; Ohigashi, H.; Osawa, T & Uchida. K. (2003). A phase 11 detoxification enzyme inducer from lemongrass: identification of citral.
 23. Onawunmi G.O. Evaluation of the antimicrobial activity of citral. *Lett Appl Microbiol* 1989;9:105-8.
 24. Paranagama P.A., Abeysekera K.H.T., Abeywickrama K., Nugaliyadde L. Fungicidal and anti-aflatoxinigenic effects of the essential oil of *Cymbopogon citratus* (DC.) Stapf (lemongrass) against *Aspergillus flavus* Link. Isolated from stores rice. *Lett Appl Microbiol* 2003; 37:86-90.
 25. Rajesh .V, Perumal. P, Chinthakindhi .V, Prabhakaran .S, Hymavathi G, Guntupalli. T: In-Vitro Evaluation Of *Smilax Zeylanica* Linn. Leaves for Anthelmintic Activity. *The Internet Journal of Pharmacology*. 2010 Volume 9 Number 1.
 26. Rios, J.L & Recio, M., C. (2005). Medicinal plants and antimicrobial activity. *Journal Of Ethnopharmacology* 100: 80-84.
 27. Satish B. Kosalge and Ravindra A. Fursule. *Asian Journal of Pharmaceutical and Clinical Research*. Investigation of In Vitro Anthelmintic Activity of *Thespesia Lampas* (Cav.). *Volume 2, Issue 2, April- June, 2009*.
 28. Schuck V.J.A., Fratini M., Rauber C.S., et al. Avaliação da atividade antimicrobiana de *Cymbopogon citratus*. *Revista Brasileira de Ciências Farmacêuticas* 2001;37:45-9.
 29. Shivkar YM, Kumar VL. Anthelmintic activity of latex of *Calotropis procera*. *Pharma Bio*, 2003; 41:263-
 30. Sollmann T. Anthelmintics: Their efficiency as tested on earthworms. *J Pharmacol Exp Ther* 1918; 12:129-70.
 31. Sherwani S.K, Gilani A. S, Masroor. S, Kazmi.S.U. (2013). In vitro anthelmintic activity of crude leaf extract of *Bougainvillea spectabilis*. *FUUAST J. Biol* (Accepted)
 32. Sherwani S.K, Khan M. M, Munir S., Shah M.A., Kazmi S.U. Anthelmintic potential of crude extract of *Camellia sinensis* (Green tea). *International Research Journal of Pharmacy*(2013) (Accepted).
 33. Szewezuk VD, Mongelli ER, Pomilio AB. Antiparasitic activity of *Melia azadirach* growing in Argentina. *Molecular Med Chem* 2003; 1:54-7.
 34. Thorn GW, Adams RD, Braunwald E, Isselbacher KJ, Petersdorf RG. *Harrison's Principles of Internal Medicine*. New York: McGraw Hill Co; 1977.
 35. Trapti R, Vijay B, Komal M, Aswar PB, Khadbadi SS: Comparative studies on antihelmintic activity of *Moringa oleifera* and *Vitex negundu*, *Asian J. Research Chem.*, 2(2), 2009:181-182
 36. Vidyarthi RD. *A Text Book of Zoology*. 14th ed. New Delhi: S. Chand and Co; 1967.
 37. Vidyadhar.S, Saidulu. M, Gopal T.K, Chamundeewari. D, Rao.U, Banji.D. *In Vitro* anthelmintic activity of the whole plant of *Enicostemma littorale* by using various extracts .Volume: I: Issue-3: Nov-Dec -2010
 38. Vigar Z. *Atlas of Medical Parasitology*. 2nd ed. Singapore: P.G. Publishing House; 1984.
 39. Wannissorn B., Jarikasem S., Soontorntanasart T. Antifungal activity of lemon grass and lemon grass oil cream. *Phytotherapy Res* 1996; 10:551-4.
 40. Wannissorn B., Jarikasem S., Siriwangethachai, T., Thubthimthed, S. Antibacterial properties

of essential oils from Thain medicinal plants.
 Fitoterapia 2005; 76:233-6.

41. Zafar I, Qazi KN, Khan MN, Akhtar MS, Faisal NW, *In vitro* anthelmintic activity of *Allium sativum* *Zingiber officinale*, *Cucurbita*

mexicana, *Ficus religiosa*, International Journal of Agriculture and Biology, 3(4),2001: 454-457

Table 1: Anthelmintic activity of control and standard

Concentration mg/ml	Control		Standard	
	Paralysis time (Mins)	Death time (Mins)	Paralysis time (Mins)	Death time (Mins)
25mg/ml	96.46 ± 0.46	160.06±0.11	18.36 ± 0.15	54.36 ± 0.40
50mg/ml	78.23 ± 0.25	142.2±0.34	12± 0.00	46.46 ±0.40
100mg/ml	62.43±0.11	111.7±0.26	08.84 ± 0.03	13.6±0.17

Values are the mean ± S.E.M. of control and standard drug on three earthworms. Control is normal saline while standard drug is piperzine citrate

Table 2: Anthelmintic activity of crude extract of *Cymbopogon citratus* (Lemon grass)

Concentration mg/ml	Crude extract of <i>Cymbopogo citratus</i> (lemon grass)	
	Paralysis time (mins)	Death time (mins)
25mg/ml	44.86± 0.41	120.46 ±0.25
50mg/ml	22.16± 1.06	92±0.51
100mg/ml	10.7 ± 0.43	80.7±0.51

Values are the mean ± S.E.M. of aqueous extract of *Cymbopogon citratus* (Lemon grass) on three earthworms